Response of 3D tumors to Photodynamic Therapy evaluated with Raman microspectroscopy

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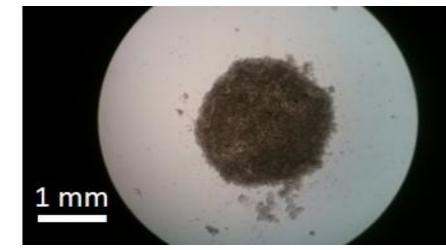
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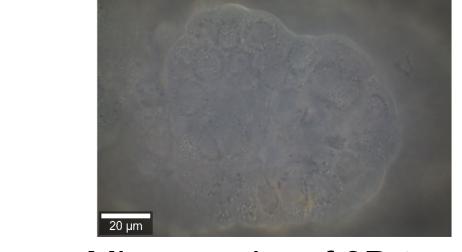
Introduction

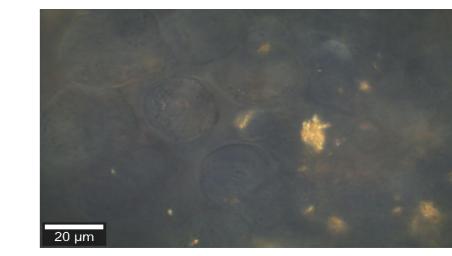
Photodynamic Therapy is a technique used as an alternative for the treatment of various types of cancer in the clinic, including the breast cancer. It is important to understand the effects of PDT on solid tumors, since many studies are done on 2D culture and the 3D culture more closely resembles the natural tissue.

The purpose of this project is to evaluate the effects of Photodinamicy Terapy (PDT) in 3D cultures of tumor cells using the Raman microspectroscopy.

Results



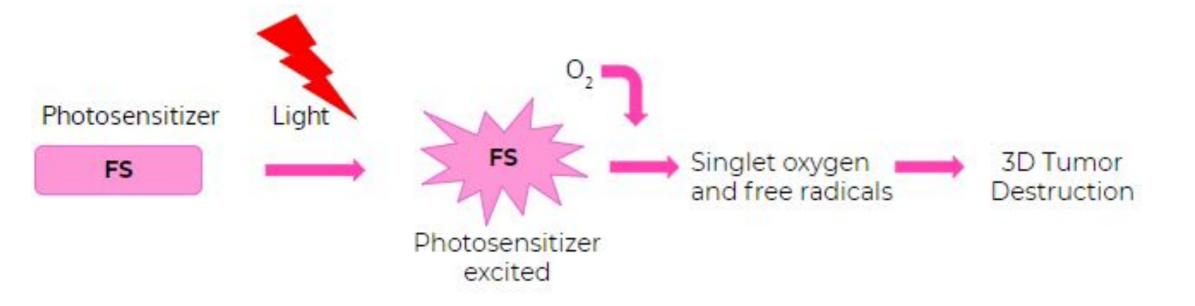




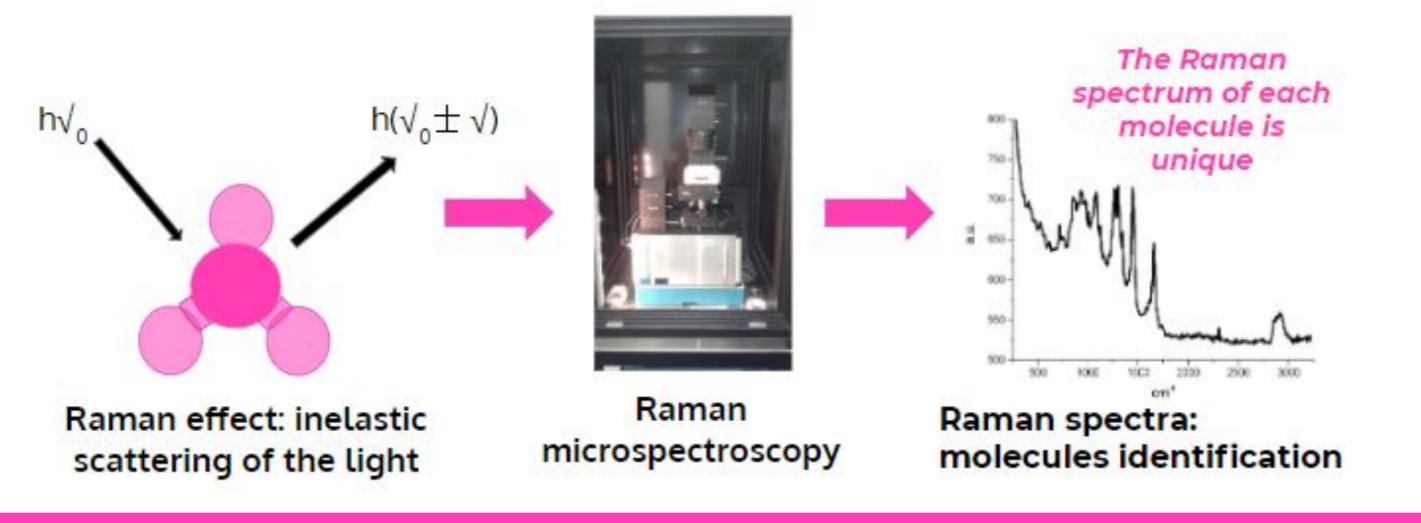
3D tumor after 4 days of growing.

Micrographs of 3D tumors before (left) and after (right) PDT.

Magnetic nanoparticles, used to form the tumor, work as a SERS nanoparticles in Raman microspectroscopy of spheroids and help to enhance the Raman signal from the cells.

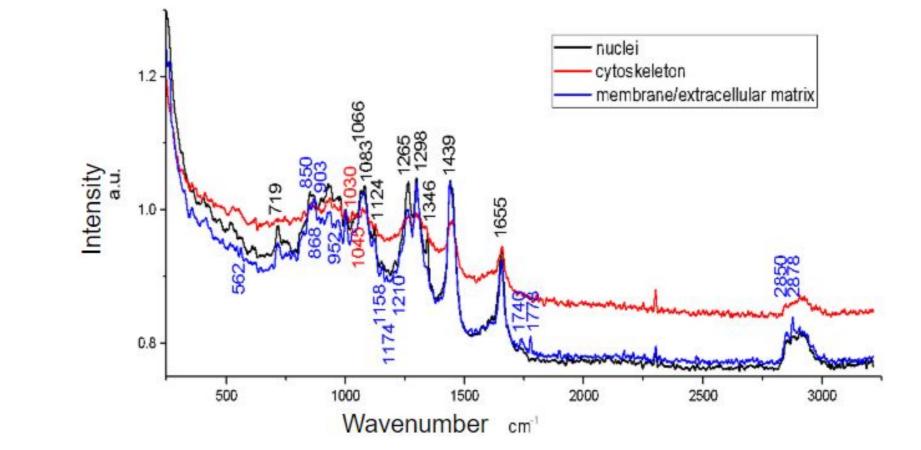


microspectroscopy is one of the most suitable Raman methods for discovery molecular mechanisms of the metabolic process and it is possible to study live cell samples without the use of markers.



Materials and Methods





The membrane/ ECM spectrum:

glycine (562 and 1158 cm⁻¹) [1] proline (850, 903, 952 and 1174 cm⁻¹) [1] \longrightarrow Extracellular matrix fibronectin (1740 cm⁻¹) [3]

After PDT

lipids (2850 e 2878 cm⁻¹) [1] Membrane

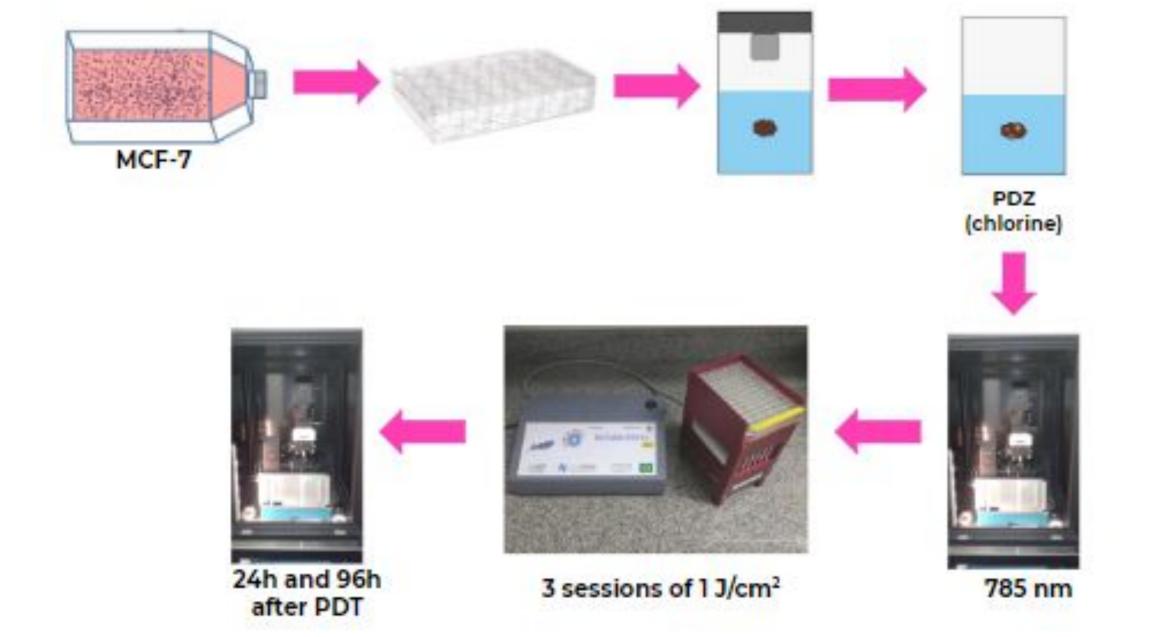
24h after PDT:

decrease in spectra:

The nuclei spectrum: nucleic acids (719, 1124, 1265, 1346, 1439 and 1655 cm^{-1}) [1]

The cytoskeleton

spectrum: the bands shape the and of cytoskeleton spectrum in the study of Klein and cols [2]



Acknowledgements



References

[1] Gelder J. D. et al. Reference database of Raman spectra of biological molecules. J. Raman Spectrosc. 2007; 38: 1133–1147. [2] Klein, K. et al. Label-Free Live-Cell Imaging with Confocal Raman Microscopy. Biophysical Journal V. 102, 2012, pp. 360–368. [3] Strehle M. A. A Raman spectroscopic study of the adsorption of fibronectin and fibrinogen on titanium dioxide nanoparticles. Phys. Chem. Chem. Phys., 2004, 6, 5232 - 5236.

[4] Nguyen T. T. et al. Characterization of Type I and IV Collagens by Raman Microspectroscopy: Identification of Spectral Markers

• Lipids (2850-2880 cm⁻¹) [4], nucleic acids (1265, 1439 and 1655 cm^{-1}) [5]

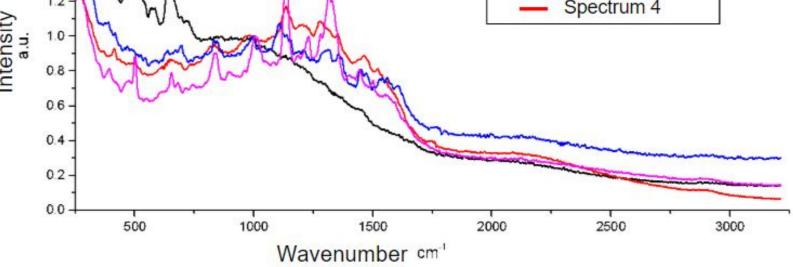
increase in spectra:

• S-S bond of fibronectin $(502-506 \text{ cm}^{-1})$ [4], [6]

96 hours after the PDT:

Increase of peaks associated with aromatic amino acids:

- Phenylalanine (363, 951, 1156 and 1351 cm⁻¹) [1]
- Tyrosine (431 and 1214 cm⁻¹)
- Tryptophan (594, 768, 1102, 1278, 1310 and 1486 cm⁻¹) [1]

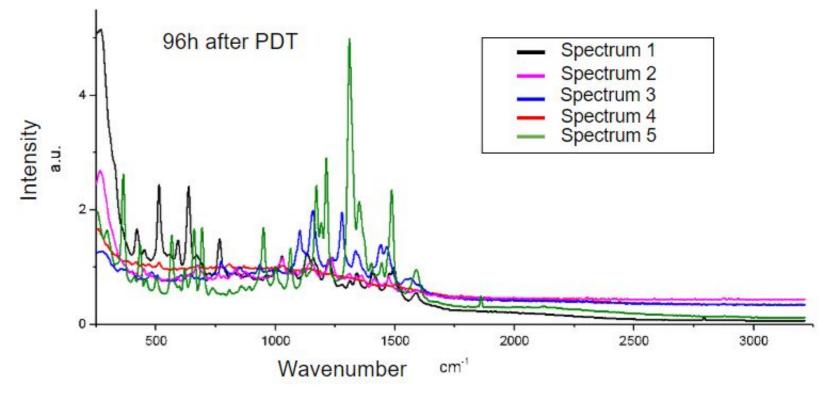


Spectrum Spectrum 2

— Spectrum 3

24h after PDT

Disaggregation of ECM and cellular membrane, exposing them the to environment.



Conclusion

With Raman microspectroscopy it is possible to distinguish cellular components and monitor changes in cells after Photodynamic Therapy in 3D culture models. In our study we

show that both tumor cells and extracellular matrix are damaged

of the Dermo-Epidermal Junction. Hindawi Publishing Corporation Spectroscopy: An International Journal Volume 27 (2012),

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[6] Rygula A. et al. Raman spectroscopy of proteins: a review. J. Raman Spectrosc. 2013, 44, 1061–1076.

